# MEDICAL AND BIOLOGICAL ASPECTS RELATED TO ASSESSMENT OF IMPACTS EXERTED BY RISK FACTORS

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## STUDYING THE CONTAMINATION OF TEA AND HERBAL INFUSIONS WITH MOLD FUNGI AS POTENTIAL MYKOTOXIN PRODUCERS: THE FIRST STEP TO RISK ASSESSMENT (MESSAGE 1)

# L.P. Minaeva, A.I. Aleshkina, Y.M. Markova, A.S. Polyanina, T.V. Pichugina, I.B. Bykova, V.V. Stetsenko, N.R. Efimochkina, S.A. Sheveleva

Federal Research Centre for Nutrition, Biotechnology and Food Safety, 2/14 Ust`inskiy pr., Moscow, 109240, Russian Federation

We analyzed microbe contamination of 54 tea samples (Camellia sp.), black and green one, including those with various additives, and tea infusions, including herbal ones. Tea that was not packed (semi-finished product) came from the following regions: India, Indonesia, Sri-Lanka, Vietnam, Kenya, China; packed tea was bought in retail outlets in the RF. Overall, 83.3 % samples of unpacked tea conformed to microbiological standards as per mold fungi; 16.7 % samples that didn't conform to them contained mold fungi in quantities equal to  $1.3-8.2\cdot10^3$  CFU/g. We detected discrepancies in quantities of mold fungi in samples with different fraction structure of tea (in average CFU/g): large-leaved tea contained  $2,3\cdot10^2$  CFU/g; middle-leaved,  $7,4x10^2$ ; small-leaved (including tea dust),  $1,7\cdot10^3$ . All packed tea samples (Camellia sp.), including those with additives, conformed to the requirements fixed by the existing standards. Aspergillus niger mold fungi prevailed in examined tea (Camellia sp.). We revealed substantial microbe contamination in herbal teas; 55 % samples didn't conform to the existing standards and contained more than  $10^{4-6}$  CFU/g of mold fungi. Besides, 72.2 % of these samples contained more than  $10^{5-8}$  CFU/g of bacteria; 62.5 % samples of herbal teas that conformed to the standards were contaminated with great quantities of bacteria equal to  $8\cdot10^5 - 2\cdot10^8$  CFU/g. We detected Aspergillus, Penicillium, Alternaria, Fusarium in herbal teas microflora; they were producers of hazardous mycotoxins, including emergent ones, and it could potentially cause contamination in future to identify hazards caused by mycotoxic fungi in tea and tea infusions as well as to update existing standards.

Key words: tea, Camellia sp., herbal tea, mold fungi, producers of mycotoxins, producers of emergent mycotoxins, mycotoxins, microbe contamination, bacteria.

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Liudmila P. Minaeva – Candidate of Technical Sciences, Senior Researcher at the Laboratory for biologicalsafety and nu-trimicrobiom analysis (e-mail: liuminaeva-ion@mail.ru; tel. 8 (495) 698-53-83; ORCID: http://orcid.org/0000-0003-1853-5735).

Anastasiya I. Aleshkina – Junior research worker at the Laboratory of biological safety and nutrimicrobiom analysis (e-mail: ninecatlifes@yandex.ru; tel. 8 (495) 698-53-83; ORCID: https://orcid.org/0000-0001-5010-2038).

Yulia M. Markova – Candidate of Biological sciences, Research worker at the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: yulia.markova.ion@gmail.com; tel. 8 (495) 698-53-83; ORCID: http://orcid.org/0000-0002-2631-6412).

Anna S. Polyanina – Research assistant at the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: polyanina.anna.sergeevna@gmail.com; tel. 8 (495) 698-53-83; ORCID: https://orcid.org/0000-0002-2766-7716).

Tatiana V. Pichugina – Candidate of Technical Sciences, Researcher at the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: bbtvp@ion.ru; tel. 8 (495) 698-53-83; ORCID: http://orcid.org/0000-0002-4632-7119).

Irina B. Bykova – Researcher at the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: bikova@ion.ru; tel. 8 (495) 698-53-83; ORCID: https://orcid.org/0000-0001-7288-312X).

Valentina V. Stetsenko – Junior researcher at the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: stetsenko\_valentina1992@mail.ru; tel. 8 (495) 698-53-83; ORCID: http://orcid.org/0000-0001-6470-171X).

Natalia R. Efimochkina – Doctor of Biological Sciences, Leading researcher at the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: karlikanova@ion.ru; tel. 8 (495) 698-53-83; ORCID: http://orcid.org/0000-0002-9071-0326).

Svetlana A. Sheveleva – Doctor of Biological Sciences, head of the Laboratory for biological safety and nutrimicrobiom analysis (e-mail: sheveleva@ion.ru; tel. 8 (495) 698-53-83; ORCID: https://orcid.org/0000-0001-5647-9709).

Microscopic mold fungi are global contaminants polluting food vegetable raw materials; their prevalence is primarily determined by natural factors, and only to a certain extent, by anthropogenic influence. They can penetrate vegetable raw materials at all the stages including vegetation, picking, transportation, and storage. Certain species of mold fungi are potentially able to synthesize mycotoxins (MT) as secondary metabolites, and special attention should be paid to their producers as they contaminate food products. The most significant fungi here belong to the following stems: Aspergillus, Penicillium, Fusarium, Alternaria and Claviceps [1]. A MT range produced by these microscopic fungi includes both highly toxic ones such as zearalenone (ZEA), deoxynivalenol (DON), aflatoxins (AF B1), fumonisins (FB1 and FB2), ochratoxin A (OTA) and patulin, T-2 toxin, with their contents in food products being strictly standardized in the Customs Union countries (CU TR 021/2011)<sup>1</sup> and abroad [2-8], and emergent mycotoxins (EMT) that have not been studied enough: enniatins, bovericin, moniliformin, fuzproliferin, fusaric acid, sterigmatocystin, emodin, mycophenolic acid, alternariol and its monomethyl ether, tenuazonic acid, asperglaucid, and tentoxin, that can simultaneously occur in food products and make its toxic contribution into an end product [9-12]. Mycotoxins cause actual hazards for people as they exert carcinogenic, mutagenic, and teratogenic impacts, suppress the immune system, and can induce a number of diseases [13].

Bearing safety in mind, it is important to examine contamination with mold fungi, including EMT producers, that occurs in vegetable food products consumed by population in large volumes, for example, tea. As per data obtained in marketing research conducted in 2003 and 2018, 82-83% respondents drank tea every day [14, 15]. As there is a great variety of tea types distributed on the consumer market, such concepts as "tea" and "tea drinks" are often mixed up. According to the State Stan-

dard  $(GOST)^2$  "tea" is made of tea leaves from *Camellia* stem plants, *Theaceae* family (*Camelliasp.*) and doesn't contain any other components; "tea with additives" should contain not less than 50% tea as well as include some other components; "tea drink" is made of vegetable raw materials with probably added tea (mass fraction is not more than 50%) and other components, and this concept covers herbals teas as well. Tea leaves that were at primary processing stages, including drying, are a semi-finished product.

As pace of life is becoming faster, people are switching from leaf teas to packed ones (tea in bags for brewing); thus, as per market analysis data obtained in 2016, packed teas had about 63% share of the overall consumption [18]. There are also changes in preferences of specific consumer groups as mature and elderly people tend to choose green or fruit tea; as per data collected in 2016 green tea had 23% share of the market [15], fruit and herbal teas, 3.5-5%, and these market segments were considered to be growing [15-16].

Most people tend to think that tea is a "useful", "healthy" and "safe" drink and it is due to both heat treatment involved in the process of making it and absence of any relations with food intoxications. CU TR fixes microbiological standards for tea and tea products that are to be met in the RF; these parameters are given in Table 1.

Concentration of mycotoxic aflatoxin B1 produced by *A. flavus, A. parasiticus, A. ochraceoroseus, A. pseudotamarii, A. bombycis, A. nomius* is regulated in tea and tea products and it shouldn't exceed 0.005 mg/kg [17].

Safety parameters regulated and standardized in other countries are determined by national legislation and can include either a single parameter or a whole set of them. Parameters recommended by Tea & Herbal Infusions Europe (THIE, 2018) [18-19], American Herbal Products Association (AHPA, 2016) [20] and by the WHO (2007) [21] are given in Table 2.

<sup>&</sup>lt;sup>1</sup> The Customs Union Technical Regulations CU TR 021/2011 "On food products safety".

<sup>&</sup>lt;sup>2</sup>GOST 32593-2013 Tea and tea products. Terms and definitions.

## Table 1

Microbiological safety parameters for tea and tea products according to the requirements fixed in the CU TR 021/2011

A product	Amounts of mesophilic aerobic and facultatively anaerobic mi- croorganisms (AMAFAnM)	Mold	Yeast	B.cereus	Coliform bacteria	E. coli	S.aureus	Pathogenic bacteria including salmonella
	CFU	/g, not	more				a product,	g, not allowed
Теа		10 <sup>3</sup>	_	-	-	-		-
Mixtures of dried vege- table raw materials for making hot non- alcoholic drinks	5x10 <sup>5</sup>	100	100	-	1	-	-	25
BAD to food - mix- tures of dried medicinal herbs (teas)	5x10 <sup>5</sup>	10 <sup>3</sup>	100	-	0.01	0.1	-	10
BAD-teas (dried mix- tures for children)	5x10 <sup>3</sup>	50	50	200	0.1	1	1	25
<ul> <li>Herbal instant teas (made of plants) for pregnant and breast- feeding women;</li> <li>Herbal drink for chil- dren (herbal tea)</li> </ul>	5x10 <sup>3</sup>	50	50	100	1	-	-	25

## Table 2

Microbiological parameters for tea, herbal drinks, and raw materials

A product	Aerobic bacteria quantity, CFU/g	Yeast, CFU/g	Mold, CFU/g	Enterobac- teriaceae, CFU/g	<i>E. coli</i> , CFU/g	<i>Salmonella</i> , absence in a mass of product, g
Tea	& Herbal Infu			<u>E)</u>		
Tea (C. sinensis)	$\leq 10^7$	$\leq 10^{6}$	$\leq 10^5$	-	$\leq 10^2$	125
Raw materials for herbal teas (dried)	$\leq 10^8$	$\leq 10^{6}$	$\leq 10^{6}$	-	$\leq 10^4$	125
Herbal teas (dried)	$\leq 10^7$	$\leq 10^5$	$\leq 10^5$	-	$\leq 10^3$	125
Drinks made of tea ( <i>C. sinensis</i> ) and herbal teas (cooled)	$\leq 10^4$	$\leq 10^2$	$\leq 10^2$	-	$\leq 10^1$	125
America	n Herbal Proa	lucts Assoc	ciation (A	HPA)		
Herbal additives (raw materials)	$\leq 10^7$	$\leq 10^5$ ,	totally	$\leq 10^4$	≤10	25
И	orld Health C	Prganizatio	on (BO3)			
Herbal medications taken after	$\leq 10^7$	$\leq 10^4$ ,	totally	$\leq 10^3$	10	1
being brewed with boiling water	Not allowed in 1 g, Clostridia and Shigella					
Vegetable materials to be con-	$\leq 10^5$ $\leq 10^3$ , totally		$\leq 10^3$	10	1	
sumed as food	Not allowed in	n 1 g, <i>Clos</i>	<i>tridia</i> and	Shigella		

But still, there are no unified criteria in the international legislation. Low humidity of tea prevents microbe contamination development; however, when requirements to picking and processing are violated, or cross contamination occurs at a manufacturing plant, it can cause elevated risks of finished products being contaminated, and it is particularly true for herbal teas.

Research on microbe contamination that can be found in foreign literature primarily focuses on teas that underwent deep fermentation, for example, Pu'er teas; but such teas have rather insignificant share on the market, and as for mass consumption teas, there is very little research on them or such research is fragmentary [22-24]. The report on the results of the 23rd session held by the FAO Intergovernmental Group (IGG) on Tea in Hangzhou, China, on May 17-20, 2018 which was presented to the FAO Committee on Commodity Problems highlighted there was a necessity to promote scientific research in the sphere of natural contaminants and to reveal relationships that caused their occurrence in tea [25].

At present experts are being discussing an issue on microbiological parameters fixed for tea in the RF and possible alterations to them in order to harmonize the RF national legislation with international requirements recommended by the WHO. Given that, it seems vital to examine contamination of variable teas and tea products distributed in the Customs Union countries with mold fungi which are potential MT producers (including EMT); it is also necessary to assess existing microbiological standards as regards their efficiency for providing safety of tea and tea products for population.

**Our research goal** was to examine contamination of variable teas and tea products with mold fungi – mycotoxins producers, and also to determine bacterial contamination for further risk assessment and updating of hygienic standards existing for these products.

Data and methods. We performed experimental research on 54 samples that in-

cluded: unpacked traditional tea (Camelliasp.) (semi-finished product), green and black, totally 30 samples from 6 tea-producing regions (Vietnam, India, Indonesia, Kenya, China, Sri-Lanka); packed tea divided into three groups: traditional tea (Camelliasp.), tea with additives, 3 samples of each, and herbal teas, 18 samples overall (monocomponent ones: Sudan rose, fermented sallybloom, thyme, mint, camomile; and multicomponent ones containing: echinacea, origanum, brandy mint, nettle, thyme, camomile, wild rose, sage, violet, licorice, St. John's wort, melilot, inula, hawthorn, everlasting, tansy, holy-thistle, marigold, sarcarolla, birch leaves, fermented sally-bloom etc.). Unpacked tea samples were provided by wholesale companies, and packed tea samples were bought in retail outlets. Prior to microbiological research all samples were kept unopened under room temperature.

Mycological inoculation was accomplished according to the State Standard GOST  $10444.12-2013^3$ , we applied microscopic fungi identifiers for taxonomic identification [27-29], and determined amounts of mesophilic aerobic and facultatively anaerobic microorganisms according to the State Standard GOST  $10444.15-94^4$ .

**Results.** Safety of food products for consumers is provided by compliance with the fixed standards, and as data on new hazards and risk factors are accumulated or existing hazards are reduced somehow, these standards are to be updated. We examined microbe contamination of tea and tea products focusing on mold fungi as well as bacteria amounts of which in products allow to judge whether a production is hygienically clean or not.

**Determining contamination of tea samples with mold fungi.** The results of our research on contamination of unpacked tea (*Camellia sp.*) with mold fungi in 30 samples as per regions of origin are given in Table 3.

<sup>&</sup>lt;sup>3</sup> GOST 10444.12-2013 Microbiology of food products and animal forage. Procedures for detecting and canculating amounts of yeast and mold fungi.

<sup>&</sup>lt;sup>4</sup>GOST 10444.15-94 Food products. Procedures for determining amounts of mesophilic aerobic and facultatively anaerobic microorganisms.

#### Table 3

Dagian	Number	Mold con	tents, CFU/g	As per spec	cific ranges,	CFU/g (numbe	er of samples)	
Region of origin	of sam- ples	Average	Range of amounts	Less than 500	500-1,000	1,000-5,000	More than 5,000	
	pies						3,000	
			Blac	k tea				
Vietnam	5	56	18-150	5	-	-	-	
India	5	1,577	173-5,850	3	-	1	1	
Indonesia	5	2,364	110-8,250	2	1	1	1	
Kenya	5	87	11-340	5	-	-	-	
Sri-Lanka	5	609	20-2,750	4		1		
(Ceylon)	5	009	20-2,750	4	-	1	-	
China	1	600	600	-	1	-	-	
Green tea								
China	4	201	28-710	3	1	-	-	
Overall, abs.	30	-	-	22	3	3	2	
Overall, %	100	-	-	73.3%	10%	10%	6.7%	

Distribution of unpacked tea samples as per levels of contamination with mold fungi obtained via mycological analysis

As we can see from the Table 3, contents of mold fungi in the examined samples of unpacked tea varied from 11 to 8,250 CFU/g in black tea and from 28 to 710 CFU/g in green tea. Standards were violated in 5 out of 26 (16.7%) black tea samples. As we examined all the samples as per regions of origin, we revealed that all the samples from Vietnam, Kenya, and China, as well as 4 samples from Sri-Lanka, 3 from Indonesia and 3 from India corresponded to the fixed standards but at the same time some samples from India and Indonesia (2 out of 5 for each country) and Sri-Lanka (1 out of 5) contained mold fungi in quantities higher than the standards. Therefore, most samples (83.3%) of unpacked tea (Camellia sp.) corresponded to the existing standards and mold fungi contents in them didn't exceed  $10^3$  CFU/g.

As we compared mold fungi contents in samples divided as per their fractional structure according to the State Standards GOST ISO 11286-2014<sup>5</sup>, GOST 32573-2013<sup>6</sup>, and GOST 32574-2013<sup>7</sup>, average data were distributed as follows: large-leaved teas (12 samples) contained  $1.6 \times 10^2$  CFU/g; average-leaved teas

(8 samples),  $8.8 \times 10^2$  CFU/g; small-leaved teas (including tea dust) (10 samples),  $1.7 \times 10^3$  CFU/g, and it means that small-leaved tea (including tea dust) that is usually used to make granulated and packed tea is the most contaminated one with mold fungi contents in it exceeding the fixed standards.

Having examined mold fungi species in tea inoculations, we revealed that *Aspergillus niger* prevailed and it was quite characteristic for traditional tea (*Camellia sp.*) [24].

Results of mycological inoculations obtained for packed tea samples are given in Table 4.

All the examined samples of packed traditional tea (*Camellia sp.*), including tea with additives corresponded to the standards as mold fungi contents in them didn't exceed  $10^3$  CFU/g.

But only 8 out of 18 (44.4%) herbal teas met the fixed requirements as mold fungi contents in the remaining ten samples exceeded  $5x10^3$  and even reached  $10^6$  CFU/g. Species identification revealed fungi from more than 5 stems in all the herbal tea samples including *Aspergillus sp, Penicillium sp., Alternaria sp., Fusarium sp., Cladosporium sp.* etc.; certain species from these stems produce toxins. We

<sup>&</sup>lt;sup>5</sup>GOST ISO 11286-2014 Tea. Classification as per tea leaves sizes performed with grain-size analysis.

<sup>&</sup>lt;sup>6</sup>GOST 32573-2013 Black tea. Specification.

<sup>&</sup>lt;sup>7</sup> FOCT 32574-2013 Green tea. Specification.

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Packed tea	Number of sam-	Mold co	ontents, CFU/g	As per specific ranges, CFU/g (number of samples)			
	ples	Average	Range of amounts	< 500	500-1000	1000-5000	> 5000
Traditional (black and green)	3	$8.6  ext{ x10}^2$	180 - 2200	2	1	-	-
With additives (black)	3	3x10 <sup>2</sup>	< 5 - 900	2	1	-	-
Herbal	18	$9.7  ext{ x10}^4$	$< 5 - 10^{6}$	7	1	-	10

Distribution of packed tea samples as per levels of contamination with mold fungi obtained via mycological analysis

should note that such a variable microflora is specific for wild meadow herbs [29]. We performed preliminary analysis of toxins formation *in vitro* by mold contaminants extracted from several examined samples of herbal teas; the results indicated these contaminants were able to synthesize fumonisin B1. Occurrence of fumonisin B1 in herbal teas and medicinal plants was detected in Turkey [30]. Extended results on toxin formation by extracted fungi strains will be obtained in subsequent research.

Determining bacterial contamination of tea. Spore-forming bacteria are natural antagonists of mold fungi and their ratio can determine microbe contamination of vegetable raw materials. Mycological analysis of herbal teas allowed to reveal that bacteria were growing actively in spite of antibiotics contents in a nutrient medium during mycological analysis. To assess hygienic state of variable tea and tea products, we analyzed their contamination with bacteria; results obtained for unpacked tea and given in Table 5; for packed tea, in Table 6.

Most samples of traditional packed tea, 29 out of 30 (96.7%), including both black and green tea, contained overall amounts of bacteria that varied from  $<1.5x10^2$  to  $5x10^3$  CFU/g, and only 1 sample (3.3%) which came from Sri-Lanka contained bacteria in amount hat reached  $6.7x10^3$  CFU/g.

As we can see from Table 6, amounts of bacteria were low in all the examined samples of packed traditional tea (*Camellia sp.*), namely in 5 out of 6 (83.3%), and in 2 samples

of tea with additives and didn't exceed  $5x10^2$  CFU/g; it amounted to  $5x10^4$  CFU/g only in 1 sample of tea with additives.

As for herbal teas, only 5 samples out of 18 (27.8%) contained bacteria in amounts within  $5 \times 10^2$  -  $5 \times 10^5$  CFU/g range, but most of them, 13 out of 18 (72.2%), contained bacteria in amounts higher than  $5 \times 10^5$  CFU/g and sometimes this parameter even reached  $2x10^8$ CFU/g (leaves of fermented sally-bloom). It should be noted, that 10 samples (55.5%) in this group didn't correspond to microbiological standards as per mold fungi either and were considered "dirty" and 4 out of 8 "clean" samples contained bacteria in amounts varying from  $7x10^5$  to  $2x10^8$  CFU/g. We should also note that increased bacterial contamination which was on average equal to  $4 \times 10^7$  CFU/g was detected in 5 samples of herbal teas that contained leaves of fermented sally-bloom. It was apparently due to fermentation conditions favorable for development of spore-forming bacteria which, due to their antagonistic activity, are able to suppress mold fungi.

The obtained results indicate that microbiological standards for tea fixed in the CU TR 021/2011 don't provide bacterial cleanness of herbal teas, and overall amount of detected bacteria is higher than standards fixed for products that contain vegetable raw materials (Table 1) as well as standards for overall amounts of bacteria recommended by the WHO, THIE, and AHPA (Table 2).

Bacterial contamination in all the examined tea samples with high contents of bacteria was caused by spore-forming bacilli *Bacillus sp*.

### Table 5

Distribution of unpacked tea samples as per contamination with bacteria obtained via
microbiological analysis

Region Number		AMAFAnM, CFU/g		As per specific ranges, CFU/g (number of samples)					
Region	of samples	Average	Range of amounts	$<5x10^{2}$	$>5x10^{2}-5x10^{3}$	$>5x10^{3}-5x10^{5}$	>5x10 <sup>5</sup>		
Black tea									
Vietnam	5	470	$<1.5 \times 10^{2} - 1.3 \times 10^{3}$	4	1	-	-		
India	5	232	$<1.5 \text{x} 10^2 - 8 \text{x} 10^2$	4	1	-	-		
Indonesia	5	13	$<1.5 \text{x} 10^{2}$	5	-	-	-		
Kenya	5	1530	$2x10^2 - 3.3x10^3$	2	3	-	-		
Sri-Lanka	5	1365	$<1.5 \times 10^2 - 6.7 \times 10^3$	4	-	1	-		
China	1	600	600	-	1	-	-		
Green tea									
China	4	98	$<1.5x10^{2}-2.5x10^{2}$	4	-	-	-		
Totally, abs.	30	-	-	24	5	1	0		
Totally, %	100	-	-	76.7	20	3.3	0		

#### Table 6

Distribution of unpacked tea samples as per contamination with bacteria obtained via microbiological analysis

Packed tea Number	AMA	AFAnM, CFU/g	As per specific ranges, CFU/g (number of samples)				
of samples		Average	Range of amounts	$<5x10^{2}$	$>5x10^{2}-5x10^{3}$	$>5x10^{3}-5x10^{5}$	$>5x10^{5}$
Traditional	3	77	$< 1.5 \text{x} 10^2 \text{ - } 1.8 \text{x} 10^2$	3	-	-	-
With additives	3	$1.6 \text{ x} 10^4$	$<1.5 \text{x} 10^2 \text{ - } 5 \text{ x} 10^4$	2	-	1	-
Herbal	18	$1.3 \text{ x} 10^7$	$4.7 \times 10^3 - 2 \times 10^8$	-	1	4	13

**Determining microbe contamination of tea after heat treatment.** To assess whether tea is safe when it is ready for consumption, it is necessary to examine how heat treatment during brewing influences microbe contamination of tea with mold fungi and bacteria. In order to do that, we took 6 samples with different microbe contamination (3 out of each group: traditional unpacked tea and herbal tea), poured them with boiling water (in a ratio 10 g tea per 90 ml of water), then brewed them for 10 minutes (just as any ordinary tea); infusions were then cooled and inoculated, and results recalculated as per dry tea are given in Table 7.

As we can see from the Table 7, heat treatment results in considerable fall in amounts of mold fungi in all tea samples from its initial level that was  $>10^2 - 10^6$  CFU/g to a level below test sensitivity which was < 5 CFU/g. However, we

# Table 7

Comparative analysis of mold fungi and bacteria contents in inoculations before and after heat treatment: results obtained for traditional teas and herbal teas

	Mold fung	gi, CFU/g	AMAFAnM, CFU/g					
Sample	Initial	After	Initial	After				
	product	brewing	product	brewing				
Traditional (unpacked) tea								
1	$5.8 \text{ x} 10^3$	< 5	$8 \text{ x} 10^2$	$< 1.5 \text{ x} 10^2$				
2	$8.2  ext{ x10}^3$	20	$< 1.5 \text{ x} 10^2$	$< 1.5 \text{ x} 10^2$				
3	$6 \text{ x} 10^2$	< 5	$1.3 \text{ x} 10^3$	$<1.5 \text{ x}10^{2}$				
	He	erbal (pack	ed) tea					
4	Less	<5	$2x10^{8}$	6.1 x10 <sup>4</sup>				
-	than 5		2/10	0.1 X10				
5	$10^{6}$	< 5	$5 \text{ x} 10^6$	$2.7 \text{ x} 10^5$				
6	$2.2 \text{ x} 10^4$	< 5	$9 \text{ x} 10^6$	$4 \text{ x} 10^6$				

didn't observe a similar fall as regards bacteria: their amount dropped to less than  $1.5 \times 10^2$ CFU/g in all 3 samples of traditional tea regardless of their levels in initial products while as for herbal teas, the greatest reduction, more than by 3,000 times, was detected in sample No. 4; bacteria amount dropped by 18 times in sample No. 5; and it fell by only 2.25 times in sample No. 6. And we should also note that amounts of bacteria remained rather high in all three samples of herbal tea even after heat treatment, from  $6.1 \times 10^4$  to  $4x10^{6}$  CFU/g (recalculated as per dry product). These differences in inhibition under heat treatment are apparently caused by prevalence of spore-forming bacteria which are heat-resistant over vegetative ones.

Conclusion. We analyzed microbiological state of different teas (Camelliasp.): black and green tea (packed and unpacked), including tea with additives, and revealed that most samples corresponded to microbiological standards fixed in CU TR 021/2011 as amounts of mold fungi didn't exceed it in 83.3% of unpacked tea samples (semi-finished product). All the samples of packed tea, including those with additives, contained less than  $10^3$  CFU/g. We detected some differences in mold fungi quantities in tea samples with different fractional structure (in average CFU/g): they were less than  $10^3$  in large-leaved and average-leaved teas, and  $1,7x10^3$  in small-leaved teas (including tea dust). Aspergillusniger prevailed among mold fungi detected in tea (Camelliasp.). After traditional tea (Camelliasp.) was brewed, the amount of mold fungi and bacteria in brewed drinks dropped from the initial level equal to  $10^{2-3}$  CFU/g to a level below test sensitivity and it made tea a safer drink for consumption. As we compared standards existing for tea in the Customs Union countries with standards recommended for tea (C.sinensis) by THIE association, we revealed that CU TR 021/2011 fixed stricter requirements to mold fungi contents, but a set of microbiological parameters fixed by THIE standards included 4 additional groups of microorganisms.

High microbe contamination was most frequently detected in herbal tea samples, both in terms of mold fungi and bacteria. 55.5% samples contained mold fungi in quantities higher than  $10^{4-6}$  CFU/g and it exceeded microbiological standards, and 72.2% samples contained overall quantity of bacteria that was higher than  $10^{5-8}$  CFU/g. And even when herbal tea samples corresponded to the fixed standards as per mold fungi contents, 62.5% of them still contained bacteria in quantity that was higher than  $5 \times 10^{\circ}$ CFU/g. After herbal teas were brewed, mold fungi contents went down from the initial  $10^{4-6}$  CFU/g to a level below test sensitivity, and bacteria quantity dropped from  $10^{6-8}$  CFU/g to more than  $10^{4-6}$  CFU/g, but it still remained rather high; that is, brewing doesn't provide microbiological safety of herbal tea as opposed to traditional tea (Camelliasp.).

The obtained results that allow to see hygienic state of herbal teas apparently indicate that when the existing microbiological standards for tea are applied to herbal teas, it doesn't provide their cleanness from bacteria. High bacteria quantities up to  $2x10^8$  CFU/g exceed both AMAFAnM parameter fixed in the CU TR 021-2011 for all types of products that include herbal raw materials (Table 1) and quantities of bacteria recommended by the WHO and THIE and AHPA associations (Table 2). In order to make tea products safer for consumers, it is advisable to adjust "tea" category and make it "tea (Camelliasp.)", to single out "herbal tea" as a specific food product that is different from "tea (Camelliasp.)", and to specify a group of products "herbal tea" can be assigned into taking into account a wider list of parameters to be standardized.

We detected a great variety of microscopic fungi in herbal teas including *Aspergillussp, Penicilliumsp., Alternariasp., Fusariumsp., Cladosporiumsp.*; some of them are toxins producers. It allows us to assume there are mycotoxins produced by these fungi in herbal teas. To obtain a most comprehensive characteristics of tea samples, it is necessary to analyze mycotoxins occurrence in them with a multi-detection procedure aimed at detecting a wider range of mycotoxins including emergent ones. When assessing safety of tea and tea products, it is advisable to take into account all the existing data obtained in microbiological and mycotoxicological research.

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#### References

1. Bennet, J.W.; Klich, M. Mycotoxins. *Clinical Microbiology Reviews*, 2003, no. 16, pp. 497–516 [CrossRef].

2. COMMISSION REGULATION (EC) No 1881/2006 of 19.12.2006 setting maximum levels for certain contaminants in foodstuffs. *OJ*, L 364, 20.12.2006, 5 p.

3. COMMISSION REGULATION (EU) No 165/2010 of 26.02.2010 amending Regulation (EC) No 1881/2006 setting maximum levels for certain contaminants in foodstuffs as regards aflatoxins. *OJ*, 27.2.2010, L 50, 8 p.

4. Commission Regulation (EU) No 1058/2012 of 12.11.2012 amending Regulation (EC) No 1881/2006 as regards maximum levels for aflatoxins in dried figs Text with EEA relevance. *OJ*, 13.11.2012, L 313, pp. 14–15.

5. COMMISSION REGULATION (EU) No 105/2010 of 05.02.2010 amending Regulation (EC) No 1881/2006 setting maximum levels for certain contaminants in foodstuffs as regards ochratoxin A. *OJ*, 2010, L 35, 7 p.

6. Commission Regulation (EU) 2015/1137 of 13.07.2015 amending Regulation (EC) No 1881/2006 as regards the maximum level of Ochratoxin A in Capsicum spp. spices (Text with EEA relevance). *OJ*, 14.07.2015, L 185, pp. 11–12.

7. Commission Recommendation 2013/165/EU of 27.03.2013 on the presence of T-2 and HT-2 *toxin* in cereals and cereal products Text with EEA relevance. *OJ*, 03.04.2013, L 91, pp. 12–15.

8. Commission Regulation (EC) No 1126/2007 of 28.09.2007 amending Regulation (EC) No 1881/2006 setting maximum levels for certain contaminants in foodstuffs as regards Fusarium toxins in maize and maize products (Text with EEA relevance) *OJ*, 29.09.2007, L 255, pp. 14–17.

9. Sedova I., Kiseleva M., Tutelyan V. Mycotoxins in Tea: Occurrence, Methods of Determination and Risk Evaluation. *Toxins*, 2018, no. 10, 444 p. DOI: 10.3390/toxins10110444 [CrossRef] [PubMed] (in Russian).

10. Zhang L., Dou X.W., Zhang C., Logrieco A.F., Yang M.H. A Review of Current Methods for Analysis of Mycotoxins in Herbal Medicines. *Toxins*, 2018, no. 10, 65 p. [CrossRef] [PubMed].

11. Jestoi M. Emerging fusarium-mycotoxinsfusaproliferin, beauvericin, enniatins, and moniliformin: a review. *Critical Reviews in Food Science and Nutrition*, 2008, vol. 48, no. 1, pp. 21–49. DOI: 10.1080/10408390601062021

12. Kovalsky P., Kos G, Nahrer K. [et al.]. Co-Occurrence of Regulated, Masked and Emerging Mycotoxins and Secondary Metabolites in Finished Feed and Maize – An Extensive Survey. *Toxins*, 2016, vol. 12, no. 8, 363 p. DOI: 10.3390/toxins8120363

13. Wu F., Groopman J.D., Pestka J.J. Public health impacts of foodborne mycotoxins. *Annual Review of Food Science and Technology*, 2014, no. 5, pp. 351–372 [CrossRef] [PubMed].

14. Martinchik A.N., Baturin A.K. Martinchik A.E. Potreblenie chaya i kofe naseleniem Rossii [Tea and coffee consumption by the population of Russia]. *Voprosy pitaniya*, 2005, no 3, pp. 42–46 (in Russian).

15. Marketing research of tea consumers. Marketing research agency FDFgroup. Available at: http://fdfgroup.ru/poleznaya-informatsiya/gotovye-issledovaniya/marketingovoe-issledovanie-potrebite-ley-chaya/ (14.11.2018).

16. Tea market of Russia: new opportunities in conditions New Normal. Available at: https://www.retail-loyalty.org/journal retail loyalty/read online/art197823/ (14.11.2018) (in Russian).

17. The Aspergillus Website. Available at: https://www.aspergillus.org.uk/metabolites?title=aflato-xin&field trivial name value=&field molecular weight value=&field produced by value (14.11.2018).

18. Tea & Herbal Infusions Europe (THIE). Compendium of Guidelines for Tea (Camellia sinensis). Available at: http://www.thie-online.eu/fileadmin/inhalte/Publications/Tea/2018-08-20\_Compendium\_of\_Guidelines\_for\_Tea\_ISSUE\_5.pdf (14.11.2018).

19. Tea & Herbal Infusions Europe (THIE). Compendium of Guidelines for Herbal and Fruit Infusions. Available at: http://www.thie-online.eu/fileadmin/inhalte/Publications/HFI/2018/2018-07-17\_Compendium of Guidelines for Herbal Infusions - ISSUE 6.pdf (14.11.2018).

20. American Herbal Product Association (AHPA). Recommended microbial limits for botanical ingredients (in colony-forming units (CFU)/g). 2016. Available at: http://www.ahpa.org/Portals/0/PDFs/ Policies/14 0206 AHPA micro limits comparisons.pdf (14.11.2018).

21. WHO guidelines for assessing quality of herbal medicines with reference to contaminants and residues. 1. Medicine, Herbal - standards. 2. Quality control. 3. Drug contamination. 4. Chemistry, Analytical. 5. Guidelines. I. World Health Organization, 118 p. Available at: http://apps.who.int/medicine-docs/documents/s14878e/s14878e.pdf (14.11.2018).

22. Zhang Y., Skaar I., Sulyok M., Liu X., Rao M., Taylor J.W. The Microbiome and Metabolites in Fermented Pu-erh Tea as Revealed by High-Throughput Sequencing and Quantitative Multiplex Metabolite Analysis. *PLoS ONE*, 2016, no. 11, e0157847 [CrossRef] [PubMed].

23. Haas D., Pfeifer B., Reiterich C., Partenheimer R., Reck B., Buzina W. Identification and quantification of fungi and mycotoxins from Pu-erh tea. *International Journal of Food Microbiology*, 2013, no. 166, pp. 316–322 [CrossRef] [PubMed].

24. Carraturo F., De Castro O., Troisi J., De Luca A., Masucci A., Cennamo P., Trifuoggi M., Aliberti F. Comparative assessment of the quality of commercial black and green tea using microbiology analyses. *BMC Microbiology*, 2018, no. 18, 4 p. [CrossRef] [PubMed].

25. CCP 18/INF/13 (CCP: TE 18/Report) Report of the Twenty-Third Session of the Intergovernmental Group on Tea (Hangzhou, the People's Republic of China, 17-20 May 2018). Available at: http://www.fao.org/fileadmin/user\_upload/bodies/CCP\_72/CCP72\_INF/MX217\_INF\_13/MX217\_CCP\_18\_INF\_13\_en.pdf (14.11.2018) (in Russian).

26. Bilay V.I., Kurbatskaya Z.A. The manual for Identification toxin-producing micromycetes. Kiyev, Naukova dumka Publ., 1990, 236 p. (in Russian).

27. Sutton D.A., Fothergill A.W., Rinaldi M.G; Trans. from Engl. K.L. Tarasova K.L., Kovaleva Yu.N. Edited by Dorozhkovoy I.R. Guide to Clinically Significant Fungi. Moscow, Mir Publ., 200, vol. XVI, 468 p. (in Russian).

28. Isayeva V.S., Rattel N.N., Volkova T.N. Kratkii atlas postoronnikh mikroorganizmov v pivovarennom proizvodstve [Summary Atlas of Foreign Microorganisms in the Brewing Industry]. Moscow, 1997, 95 p. (in Russian).

29. Burkin A.A., Kononenko G.P. Mycotoxin contamination of meadow grasses in European Russia. *Agricultural Biology*, 2015, vol. 50, no. 4, pp. 503–512. DOI: 10.15389/agrobiology.2015.4.503eng (in Russian).

30. Omurtag GZ, Yazicioğlu D. Determination of fumonisins B1 and B2 in herbal tea and medicinal plants in Turkey by high-performance liquid chromatography. *Journal of Food Protection*, 2004, vol. 67, no. 8, pp. 1782–1786.

Minaeva L.P., Aleshkina A.I., Markova Y.M., Polyanina A.S., Pichugina T.V., Bykova I.B., Stetsenko V.V., Efimochkina N.R., Sheveleva S.A. Studying the contamination of tea and herbal infusions with mold fungi as potential mykotoxin producers: the first step to risk assessment (message 1). Health Risk Analysis, 2019, no. 1, pp. 93–102. DOI: 10.21668/health.risk/2019.1.10.eng

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