



## ASSESSING RISKS OF ADVERSE CLINICAL COURSE AND OUTCOME OF AN INFECTIOUS DISEASE WITH MATHEMATICAL MODELING OF EXPOSURE TO ENVIRONMENTAL FACTORS ON THE EXAMPLE OF ALUMINUM OXIDE

P.V. Trusov<sup>1</sup>, N.V. Zaitseva<sup>2</sup>, V.M. Chigvintsev<sup>1,2</sup>

<sup>1</sup>Perm National Research Polytechnic University, 29 Komsomolskiy prospect, Perm, 614990, Russian Federation

<sup>2</sup>Federal Scientific Center for Medical and Preventive Health Risk Management Technologies, 82 Monastyrskaya Str., Perm, 614045, Russian Federation

---

*Our research goal was to create a mathematical model that described anti-viral immune response regulation taking into account influence exerted by exposure to variable chemical factors. We analyzed a body response to an infection taking into account innate and adaptive immunity mechanisms. This created mathematical model helps to describe spatial distribution of immune and infectious agents in various organs and systems due to allowing for time lags occurring during interactions between different components participating in the process. The mathematical model is a system of ordinary differential equations with a retarded argument; separate addends of the model describe velocity properties of the processes that produce their effects on the development of an infectious disease. We suggest an algorithm for conducting an experiment aimed at identifying certain parameters related to influence exerted by chemical factors on interaction between the neuroendocrine and immune systems. We calculated dynamics in parameters of the immune and neuroendocrine systems when a viral infection occurs under experimental exposure to aluminum oxide. The suggested approach is applied within the concept of a multi-level human body model that takes into account interactions between systems and functional state of organs that are being examined under exposure to adverse factors of variable genesis. The conducted research provides a qualitative conception about causes that explain quantitative changes in a viral agent when an immune response occurs in a body under exposure to variable factors. This approach can be applied to adjust parameters of existing population models, spread and clinical course of different infections, and to draw up a long-term forecast of an epidemiologic situation which is necessary when risks of infectious diseases are analyzed, including those occurring when a body is exposed to adverse environmental factors.*

**Key words:** mathematical model, dynamic system, viral disease, innate immunity, adaptive immunity, neuroendocrine regulation.

---

**Introduction.** Nowadays an issue of describing interrelations in adaptive systems which modify their functioning in order to preserve their optimal state under changing outer conditions is of great interest to researchers who study neuroendocrine regulation and immune mechanisms [1, 2]. Works published in the field focus on various signs of mutual regulatory influences exerted by the systems being considered on each other [3, 4]. Some

---

© Trusov P.V., Zaitseva N.V., Chigvintsev V.M., 2019

**Petr V. Trusov** – Doctor of Physical and Mathematical Sciences, Professor, Head of Mathematic Modeling of Systems and Processes Department, Chief Researcher (e-mail: [tpv@matmod.pstu.ac.ru](mailto:tpv@matmod.pstu.ac.ru); tel.: +7 (342) 239-16-07; ORCID: <https://orcid.org/0000-0001-8997-5493>).

**Nina V. Zaitseva** – Member of the Russian Academy of Sciences, Doctor of Medical Sciences, Professor, Director (e-mail: [znv@ferisk.ru](mailto:znv@ferisk.ru); tel.: +7 (342) 237-25-34; ORCID: <https://orcid.org/0000-0003-2356-1145>).

**Vladimir M. Chigvintsev** – Researcher at Mathematic Modeling of Systems and Processes Department, post-graduate student at Mathematic Modeling of Systems and Processes Department (e-mail: [cvm@ferisk.ru](mailto:cvm@ferisk.ru); tel.: +7 (342) 237-18-04; ORCID: <https://orcid.org/0000-0002-0345-3895>).

research dwells on neuroendocrine regulation of the immune system [5, 6] and controlling influence exerted by the immune system, for example, via cytokines production, both on itself and on the neuroendocrine regulatory loop [7, 8]. Most experts in the field believe neuroendocrine and immune regulatory loops are a unified "super"-regulatory meta-system [9, 10] which coordinates a complicated multi-level regulatory process in a human body. The immune system protects a body from multiple threats, including viral infections; losses occurring due to such infectious make a considerable contribution into overall damages done to population by various health disorders and are a great medical and social problem [11].

Technogenic environmental factors can cause pathomorphism and lead to deteriorated clinical course and outcome of infectious diseases [12, 13]. Technogenic processes exert influence on regulatory (immune and neuroendocrine) systems; thus, for example, it was shown [14, 15], that technogenic chemical factors exerted negative impacts on functioning of the said systems.

Observation techniques or an experimental approach with subsequent statistical processing which are conventionally applied to assess functional disorders in the immune and neuroendocrine system, in spite of all their significance, don't fully allow to analyze mechanisms and assess consequences caused by an effect occurring when functional disorders accumulate in body systems. It is due to limitations which exist in choice of representative groups, complications related to identification and detection of basic factors, and substantial material costs which are required for organizing and conducting experiments.

Mathematical modeling seems to be one of the most efficient approaches to finding an optimal strategy for examining as well as predicting clinical course of virus diseases. This approach allows to save time and resources required for solving the above-mentioned tasks. Mathematical models make

it possible to analyze influence exerted by various factors and their combinations at an individual and population level. An example of such models is mathematical prediction models which describe correlation between human health and environmental factors [16, 17, 18].

**Our research goal** was to develop approaches to assessing body responses to an infectious disease taking into account interaction between the immune and neuroendocrine system under exposure to aluminum oxide and with functional disorders accumulating in a body.

**Data and methods.** We worked out an experimental procedure for examining negative influences exerted by chemical agents on interaction between components of the immune and neuroendocrine systems. This procedure allows to determine impacts exerted by various factors on activity of immune cells that protect a body from infections.

We chose aluminum oxide as an influencing factor in this work. Technogenic pollution with metals is a widely spread problem on industrially developed territories; peculiarities related to influences exerted by such contaminants on population health are their ability to change immune cells functioning, either inhibiting or stimulating them as well as controlling proteins produced by such cells. Aluminum compounds produce inhibiting effects on functions performed by immune cells (T- and B-lymphocytes and macrophages) and on controlling proteins production as well as ratio of immune cells quantity.

Our experiment was accomplished on an undivided population of immune cells which included T-helpers (CD3/4), B-cells (CD19/22), NK-cells (CD16/56), and cytotoxic T lymphocytes (CD3/8); the population was extracted from a peripheral blood sample. We examined a combine response given by the immune cells population to impacts, but to assess functionality of each cells type, we chose a specific parameter that reflected functions of only one type of immune cells.

Influences exerted by controlling elements, such as interleukin-1, interleukin-2, or hydrocortisone, were considered to be contributors that induced (regulated) work performed by immune cells; occurrence of viruses was another contributor (we applied a solution based on hemagglutinin and concanavalin as a viral load simulator due to its ability to induce similar responses from the immune system). Changes that occurred in functional activity of the examined immune cells population were assessed as per production of specific controlling proteins.

Concentration of a viral load simulator (a universal mitogen based on hemagglutinin and concanavalin) applied in experimental research was within 10-1,000  $\mu\text{g/ml}$  range. Operating levels of interleukin-1 were determined as per data on changes in cytokine concentration in a healthy body. Normally, interleukin-1 contents should be within 0-10  $\text{pg/ml}$ . Interleukin-2 concentrations were determined as per data obtained for healthy people and existing standards; they amounted to 10  $\text{pg/ml}$  and 100  $\text{pg/ml}$  respectively. Standard hydrocortisone concentration is 138-635  $\text{nmol/l}$ . We analyzed impacts exerted by two aluminum concentrations, 0.01  $\text{mg/l}$  and 0.1  $\text{mg/l}$ , its maximum permissible concentration in water being equal to 0.2  $\text{mg/l}$ .

We determined how many experiments we required judging from the necessity to take into account impacts exerted by all the contributors, namely interleukin-1, interleukin-2, hydrocortisone, aluminum oxide, and viral load. 16 experiments are a half-replicate of a complete factor experiment for 5 factors. Below one can see the overall view of a sought function that describes impacts exerted by the considered factors on marker proteins production by cells:

$$\begin{aligned}
 y(x_i) = & b_0 + b_1x_1 + b_2x_2 + b_3x_3 + b_4x_4 + b_5x_5 + \\
 & + b_{1,4}x_1x_4 + b_{2,4}x_2x_4 + b_{3,4}x_3x_4 + b_{1,5}x_1x_5 + \\
 & + b_{2,5}x_2x_5 + b_{3,5}x_3x_5 + b_{4,5}x_4x_5 + b_{1,4,5}x_1x_4x_5 + \\
 & + b_{2,4,5}x_2x_4x_5 + b_{3,4,5}x_3x_4x_5,
 \end{aligned} \tag{1}$$

where  $y(x_i)$  is a concentration of a marker protein that characterizes functional activity of immune cells;

$b_j$  are sought coefficients of the model;

$x_1$  is a concentration of viral load simulator;

$x_2$  is interleukin-1 concentration;

$x_3$  is interleukin-2 concentration;

$x_4$  is hydrocortisone concentration;

$x_5$  aluminum oxide concentration.

All the parameters were recalculated as per their maximum value obtained during the experiment; as a result, all the variables in the mathematical model are of the same order. Maximum interferon-gamma concentration amounts to 30.62 [ $\text{pg/ml}$ ] and interleukin, 8 [ $\text{pg/ml}$ ].

We applied a structural scheme shown in Figure 1 to describe interaction between the immune and neuroendocrine systems under exposure to aluminum oxide and with functional disorders accumulating in a body. The scheme is a set of interrelated immune and neuroendocrine system elements which are the most significant components in a body response to a virus invasion. The model takes into account functional state of organs being considered.

As we describe interactions between the immune and endocrine system which are very complicated we introduce certain simplifying assumptions into the design of our model. Cells and viruses populations are assumed to be evenly spread over the epithelial layer of a target organ at any moment. We also assume that speed of changes in any variable in the model is determined by the current values of all the variables. At present we assume that the basic processes which regulate immune protection dynamics take place in three local volumes: brains (hypophysis and hypothalamus), abdominal cavity (adrenals), and a target organ. Interaction between these three local volumes occurs with a time lag.

Protection mechanisms are activated after macrophages have removed cells affected by a virus, simultaneously information molecules of (cytokine) interleukin-1 are synthesized [19].

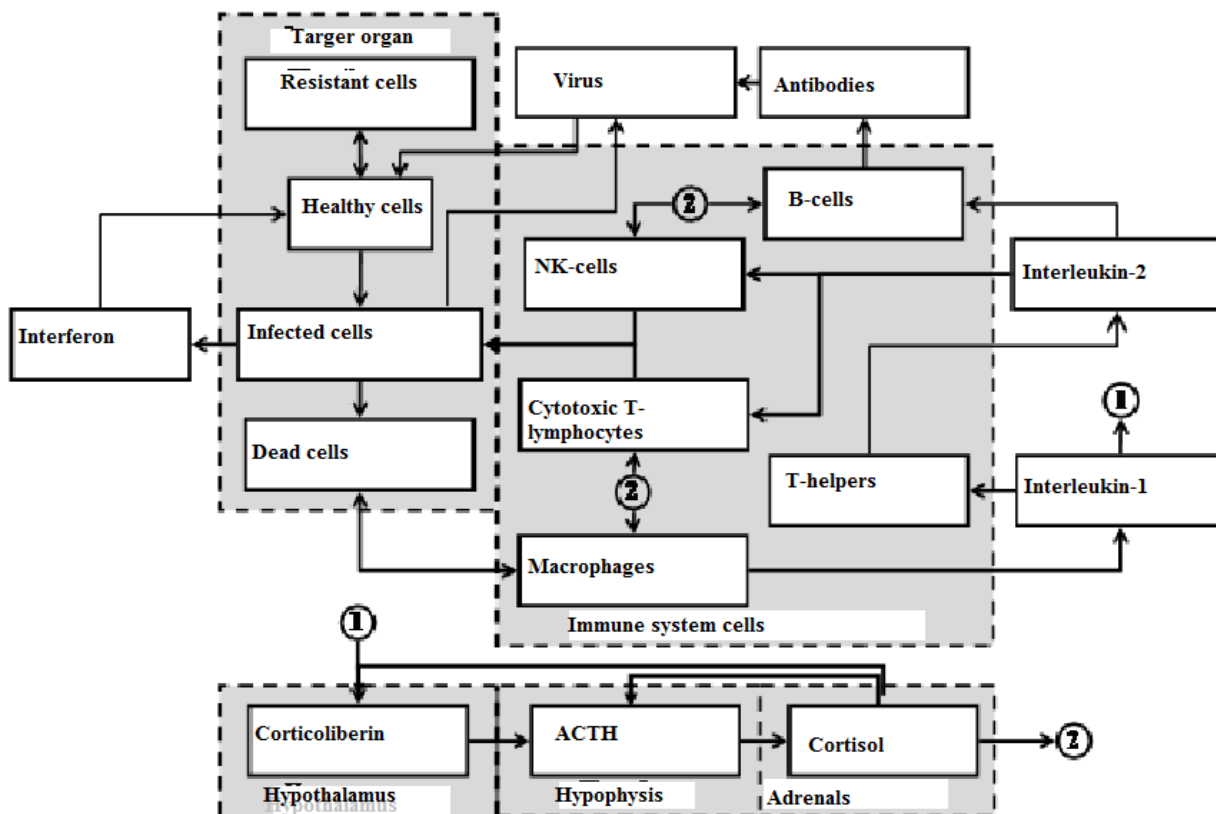


Figure 1. A conceptual scheme showing how the immune and neuroendocrine systems function in case a virus infection occurs

As interleukin-1 concentration in blood increases, it makes for T-helpers producing interleukin-2 and stimulates specific receptors in the hypothalamus to produce corticoliberin, a release hormone. Corticoliberin influences the adenohiphysis and causes adrenocorticotropic hormone (ACTH) secretion [18]. When penetrating blood, ACTH stimulates the adrenals to produce hydrocortisone; increased concentration of this hormone inhibits ACTH secretions and blocks interleukin-1 production as per negative feedback mechanism.

Regulatory impacts exerted by interleukin-2 are aimed at NK-cells [35], cytotoxic T-lymphocytes [15] and B-cells [2]. Basic NK-cells function is related to infected cells elimination at early stages of a body protecting against virus infections. In our work we allow for inhibiting effects

exerted on NK-cells by hydrocortisone [23; 24] and stimulating influence by interleukin-2 [25].

Infected cells produce interferon and it is another mechanism of primary anti-virus body protection [26, 27]. There are basic mechanisms of specific acquired immune response: B-cells produce antibodies [28] which bind free viruses, and cytotoxic T-lymphocytes destroy cells infected with viruses [29]. Hydrocortisone inhibits antiviral activity of the examined cells.

Basing on the above-given interaction scheme we can describe a mathematical model for the regulation mechanism comprising elements of the immune and endocrine system with the help of the designed model which is a system consisting of 18 ordinary first-order differential equations with a retarded argument (2):

$$\left. \begin{aligned}
 \frac{dC_{HE}}{dt} &= k_1(C_{HE} + C_R)C_D + k_2C_R - k_3C_{HE}C_{IFN} - k_4C_{HE}C_V \\
 \frac{dC_I}{dt} &= k_4C_{HE}C_V - k_5C_{NK}C_IC_{IL2} \left(1 - k_8 \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) - \\
 &- k_6C_{CTL}C_I \left(1 - k_9 \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) H(C_{CTL} - k_{46}) - k_7C_I \\
 \frac{dC_R}{dt} &= k_3C_{HE}C_{IFN} - k_2C_R \\
 \frac{dC_{IFN}}{dt} &= k_{10}C_I - k_{11}C_{HE}C_{IFN} - k_{12}C_{IFN} \\
 \frac{dC_D}{dt} &= k_7C_I + k_5C_{NK}C_IC_{IL2} \left(1 - k_8 \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) + \\
 &+ k_6C_{CTL}C_I \left(1 - k_9 \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) H(C_{CTL} - k_{46}) - k_{13}C_D C_M \\
 \frac{dC_V}{dt} &= k_{14}C_I - k_{15}C_VC_A - k_{16}C_VC_{HE} - k_{17}C_V \\
 \frac{dC_M}{dt} &= k_{18}F_b - k_{19}C_M \\
 \frac{dC_{IL1}}{dt} &= k_{20}C_M C_D \left(1 - k_{21} \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) - k_{22}C_{IL1} \\
 \frac{dC_{TH}}{dt} &= k_{23}F_b - k_{24}C_{TH} \\
 \frac{dC_{IL2}}{dt} &= k_{25}C_{TH}C_{IL1} - k_{26}C_{IL2} \\
 \frac{dC_{NK}}{dt} &= k_{27}F_b - k_{28}C_{NK} \\
 \frac{dC_{CTL}}{dt} &= k_{29} + k_{30}C_{CTL}C_{IL2} - k_{32}C_{CTL} - k_{31}C_{CTL}C_I \left(1 - k_9 \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) H(C_{CTL} - k_{46}) \\
 \frac{dC_B}{dt} &= k_{46} + k_{33}C_B C_{IL2} - k_{34}C_B \\
 \frac{dC_A}{dt} &= k_{35}C_B \left(1 - k_{36} \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) H(C_B - k_{45}) - k_{37}C_VC_A - k_{38}C_A \\
 \frac{dC_{CRH}}{dt} &= k_{48}F_h \left(1 - k_{39} \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) (1 + k_{40}C_{IL1}) - k_{41}C_{CRH} \\
 \frac{dC_{ACTH}}{dt} &= k_{49}F_p \left(1 - k_{42} \frac{C_K(t-T)}{k_{47} + C_K(t-T)}\right) C_{CRH} - k_{43}C_{ACTH} \\
 \frac{dC_K}{dt} &= k_{50}F_a C_{ACTH}(t-T) - k_{44}C_K,
 \end{aligned} \right\} \quad (2)$$

where

$C_{HE}$  is number of healthy non-resistant cells in a target organ, [cells];  
 $C_D$  is number of dead cells in a target organ, [cells];  
 $k_i$  are quotients of a model;

$C_R$  is a number of resistant cells in a target organ, [cells];  
 $C_D$  is a number of dead cells in a target organ, [cells];

$C_{IFN}$  is interferon concentration, [IU/ml];  
 $C_V$  is viruses concentration, [copies/ml];  
 $C_I$  is a number of infected cells in a target organ, [cells];

$C_{NK}$  is NK-cells (natural killers) concentration, [cells/ml];

$C_{IL2}$  is interleukin-2 concentration, [pg/ml];

$C_K$  is hydrocortisone concentration, [nanogram/ml];

$T$  is time lag, [minutes];

$C_{CTL}$  is cytotoxic T-lymphocytes concentration, [cells/ml];

$C_M$  is macrophages (monocytes) concentration, [cells/ml];

$C_A$  is antibodies concentration, [mIU/ml];

$F_b$  is marrow functional capacity, synthesizing function, [dimensionless value];

$C_{IL1}$  is interleukin-1 concentration, [pg/ml];

$C_{TH}$  is T-helpers concentration, [cells/ml];

$C_B$  is B-cells concentration, [cells/ml];

$C_{CRH}$  is corticoliberin concentration, [pg/ml];

$F_h$  is hypothalamus functional capacity, synthesizing function, [dimensionless value];

$C_{ACTH}$  is adrenocorticotropic hormone (ACTH) concentration, [picogram/ml (pg/ml)];

$F_p$  is hypophysis functional capacity, synthesizing function, [dimensionless value];

$F_a$  is adrenals functional capacity, synthesizing function, [dimensionless value].

Interactions between cells populations and body information molecules are based on the clonal selection theory (Burnet theory according to which cells clones (B-cells) occur in a body; these cells are specific to different viruses and a virus selectively contacts a corresponding clone thus stimulating it to produce antigens), mass action law (reactions speeds are proportionate to substances concentration product), interaction characteristics and Markov's death and recovery processes.

The model parameters were identified on the basis of experimental data obtained during research on a process of a body being infected with a flu virus; values for the model parameters are given in Table 1.

As all the equations in the model are complicated and non-linear, it becomes more difficult to obtain analytical solutions with it.

To solve the differential equation system, we apply implicit numerical technique by Runge-Kutta of the third order.

**Results and discussion.** We analyzed the results of our experiment during which we assessed effects produced by various factors on interferon-gamma production and determined that the greatest influence was exerted by a viral load simulator. Average standardized value for interferon-gamma amounted to  $0.225 \pm 0.058$  under low influence exerted by a viral load simulator, but when this influence was significant, it amounted to  $0.323 \pm 0.086$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ). We can conclude that interferon-gamma is synthesized due to stimulating effects produced by a viral load simulator and it is well in line with literature data.

Influence exerted by low concentrations of a viral load simulator on interferon-gamma production is significantly modified by effects produced by hydrocortisone; this phenomenon corresponds with literature data on qualitative effects produced by hydrocortisone [39]. Average standardized value for interferon-gamma concentration amounts to  $0.277 \pm 0.098$  when influence exerted by a viral load simulator is low and so is hydrocortisone concentration; but when hydrocortisone concentration is high, interferon-gamma concentration is equal to  $0.173 \pm 0.065$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ). These values are shown on Figure 2. This observed discrepancy can be an evidence that hydrocortisone has inhibiting effects under low values of a factor that influences immune cells. When a viral load simulator was high, we didn't reveal any significant effects produced by hydrocortisone.

Interferon-gamma is produced in a human body primarily by infected cells in a target organ and NK-cells. The performed experiment revealed that interferon-gamma production by NK-cells was stimulated by a controlling protein, namely interleukin-2. The obtained results are in line with literature data. Average standardized interferon-gamma concentration amounted to  $0.193 \pm 0.049$  under insignificant influence exerted by interleukin-2, but when

Table 1

Parameters of the mathematical model that describes interactions between the immune and endocrine systems under a virus infection

Parameter	Value	Source	Parameter	Value	Source
$k_1$	$2.35 \cdot 10^{-11}$ [1/cells*day]	[30]	$k_{27}$	$1.1 \cdot 10^{14}$ [cells/ml*day]	
$k_2$	0.98 [1/day]	[31]	$k_{28}$	0.11 [1/day]	
$k_3$	$1.1 \cdot 10^{-17}$ [ml/IU*day]	[32]	$k_{29}$	$4 \cdot 10^{15}$ [cells/ml*day]	[31]
$k_4$	$2 \cdot 10^{-12}$ [ml/copies*day]	[31]	$k_{30}$	4.15 [ml/pg*day]	[31]
$k_5$	$2.5 \cdot 10^{-17}$ [ml <sup>2</sup> /cells*pg*day]		$k_{31}$	$1.6 \cdot 10^{-11}$ [1/cells*day]	[32]
$k_6$	$6.6 \cdot 10^{-18}$ [ml/cells*day]	[32]	$k_{32}$	0.4 [1/day]	[31]
$k_7$	1.5 [1/day]	[33]	$k_{33}$	5.75 [ml/pg*day]	[31]
$k_8$	0.5 [dimensionless]		$k_{34}$	0.4 [1/day]	[31]
$k_9$	0.5 [dimensionless]		$k_{35}$	$7.56 \cdot 10^{12}$ [mIU/cells]	[31]
$k_{10}$	$3.2 \cdot 10^6$ [IU/cells*ml*day]		$k_{36}$	0.5 [dimensionless]	
$k_{11}$	$1.01 \cdot 10^{-10}$ [1/cells*day]	[32]	$k_{37}$	$8.6 \cdot 10^{-10}$ [ml/copies*day]	[32]
$k_{12}$	8 [1/day]	[32]	$k_{38}$	0.043 [1/day]	[31]
$k_{13}$	$10^{-14}$ [ml/cells*day]		$k_{39}$	0.5 [dimensionless]	
$k_{14}$	510 [copies/ml*cells*day]	[33]	$k_{40}$	0.002 [ml/pg]	
$k_{15}$	$8.6 \cdot 10^{-10}$ [ml/mIU*day]	[43]	$k_{41}$	3.767 [1/day]	[36]
$k_{16}$	$6.1 \cdot 10^{-12}$ [1/cells*day]	[32]	$k_{42}$	0.5 [dimensionless]	
$k_{17}$	1.7 [1/day]	[32]	$k_{43}$	0.7572 [1/day]	[37]
$k_{18}$	$3 \cdot 10^9$ [cells/ml*day]	[31]	$k_{44}$	0.1972 [1/day]	[37]
$k_{19}$	0.03 [1/day]	[31]	$k_{45}$	$1.8139 \cdot 10^{20}$ [cells/ml]	
$k_{20}$	$2.94 \cdot 10^{-19}$ [pg/cells <sup>2</sup> *say]	[34]	$k_{46}$	$0.4 \cdot 10^{16}$ [cells/ml]	[31]
$k_{21}$	0.5 {dimensionless}		$k_{47}$	3.055 [ng/ml]	[38]
$k_{22}$	0.1245 [1/day]	[35]	$k_{48}$	7.659 [pg/ml]	[36]
$k_{23}$	$5.8 \cdot 10^3$ [cells/ml*day]		$k_{49}$	21 [pg/ml]	[37]
$k_{24}$	0.0058 [1/day]		$k_{50}$	3.055 [ng/ml]	[38]
$k_{25}$	$3.28 \cdot 10^{-7}$ [ml/cells*day]		T	0.0132 [day]	[38]
$k_{26}$	0.248 [1/day]				

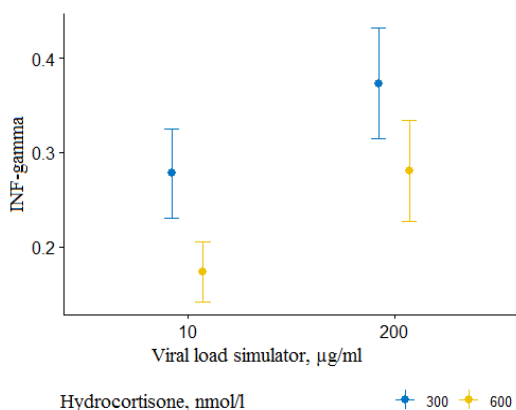


Figure 2. Average standardized interferon-gamma concentrations with error of mean under different concentration of a viral load simulator taking into account effects by hydrocortisone

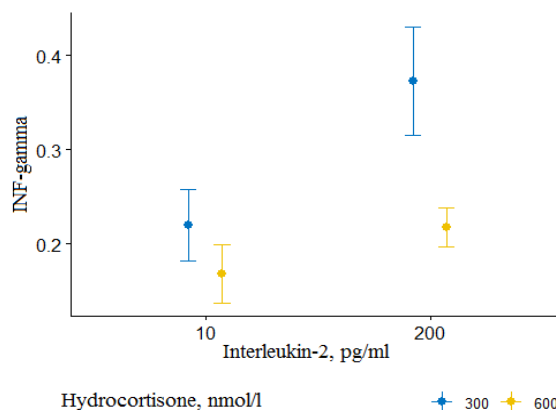


Figure 3. Average standardized interferon-gamma concentrations with error of mean under different concentration of interleukin-2 taking into account effects by hydrocortisone

this influence was high, it was equal to  $0.289 \pm 0.074$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ).

As it is not the case with a viral load simulator, hydrocortisone produces significant effects when interleukin-2 concentration is high. This observed discrepancy can be explained by the following: when we consider a viral load simulator, hydrocortisone inhibits immune reaction at the initial stage in the process thus preventing the overall chain of protective reactions in a body from activating as a virus occurs in small quantities and it is not necessary to waste energy accumulated in a body. But if interleukin-2 occurs in high concentrations in a body, it can be a sign that all the protective mechanisms are activated. In this case hydrocortisone controls interferon-gamma production in order to keep a body response to an infection within standardized limits. If the immunity exceeds these limits, interferon-gamma can damage healthy cells thus making a disease even worse and wasting greater amounts of energy.

Average standardized interferon-gamma concentration amounted to  $0.372 \pm 0.14$  under great influence exerted by interleukin-2 and low hydrocortisone concentration; but when hydrocortisone concentration was high, interferon-gamma concentration was equal to  $0.217 \pm 0.048$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ); these data are shown on Figure 3. Low hydrocortisone concentrations don't have any significant influence on interferon-gamma production stimulated by interleukin-2.

We applied the least squares method (Statistica 6.0 software) to identify parameters of the model that described effects produced by various factors (1). To test whether the model was relevant to experimental data, we applied dispersion analysis, and the obtained model had the following characteristics:  $R^2 = 0.187$  and  $p = 0.002$ . We simplified the obtained equation by dropping summands which didn't make any authentic contribution into the value for interferon-gamma concentration; this was done basing on Akaike test. We also dropped summands with low influences basing on the as-

essment of their elasticity coefficient. The equation (1), taking into account the obtained quotients values, can be given as follows:

$$y(x_i) = 0,35 + x_1(0,29x_4 - 0,32) + x_3x_5(0,72x_4 - 0,56), \quad (3)$$

where  $y(x_i)$  is standardized interferon-gamma concentration, and interferon-gamma characterizes functional activities performed by immune cells;

$x_1$  is standardized concentration of a viral load simulator;

$x_3$  is standardized interleukin-2 concentration;

$x_4$  is standardized hydrocortisone concentration;

$x_5$  is standardized aluminum oxide concentration.

The obtained relationship allows to adjust  $k_{10}$  quotient in the mathematical model (2). This adjustment allows to take into account influence exerted by aluminum oxide on an infectious process.

Figure 4 shows the results obtained via modeling three different aluminum oxide concentrations. A discrepancy in solutions to the equations systems occurs at the adaptive response stage. The obtained results revealed that exposure to aluminum oxide was at its maximum (concentration was equal to 1 mg/l), a body fought against a virus infection quite intensely but significant damage was done to it in the process. When aluminum oxide concentration was average (0.1 mg/l), dynamics of changes in flu viruses concentration corresponded to a typical average clinical course of the disease. But when exposure to an external factor was minimal (concentration was equal to 0.01 mg/l), the disease lasted longer but lesser damage was done to a body.

In case aluminum oxide significantly stimulates interferon-gamma production by NK-cells, it allows a greater number of cells in a target organ to simultaneously become resistant. It leads to a decrease in a number of healthy cells which are used by viruses for infecting. This fact makes the disease shorter but



doesn't change maximum damage done to a target organ; we chose a virus for our model identification for which the upper respiratory tracts were such an organ.

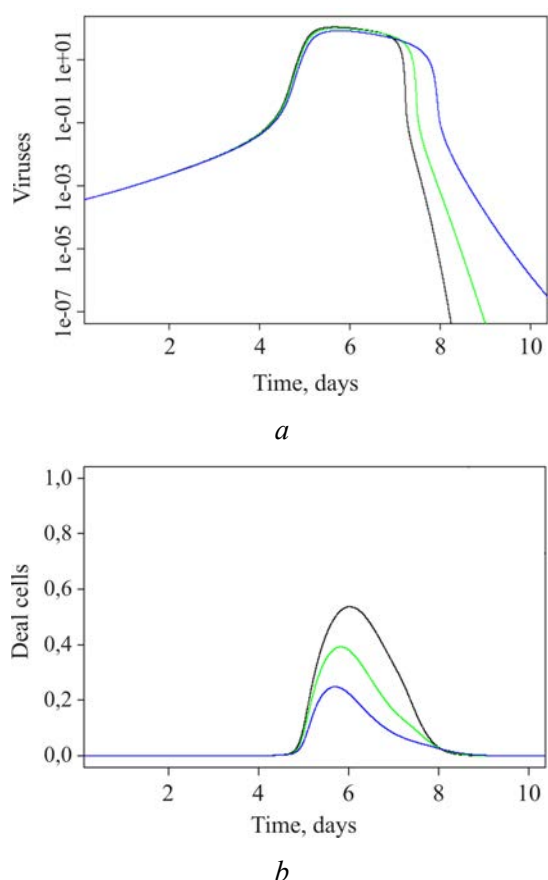


Figure 4. a) Relationship between viruses concentrations in a body and time under different aluminum concentrations b) relationship between damage to a target organ and time under different aluminum concentrations (blue line corresponds to aluminum oxide concentration equal to 0.01 ml/l; green line, 0,1 ml/l; black line, 1 mg/l)

We assessed discrepancies in average interleukin-8 concentrations under different levels of influencing factors and revealed that interleukin-1 exerted the most significant influence. Average standardized interleukin-8 concentration amounted to  $0.414 \pm 0.046$  under insignificant influence exerted by interleukin-1; but if this influence was great, it was equal to  $0.493 \pm 0.047$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ). We can conclude that interleukin-8 synthesis that characterizes NK-cells functional capabilities is stimulated by interleu-

kin-1 and it is in line with literature data [40]. Influence exerted by low interleukin-1 concentrations on interleukin-8 production is significantly modified by effects produced by hydrocortisone. This phenomenon corresponds to literature data on qualitative influence exerted by hydrocortisone on NK-cells functionality [41]. Average standardized interleukin-8 concentration amounted to  $0.448 \pm 0.08$  under insignificant influence exerted by interleukin-1 and low hydrocortisone concentration; but if hydrocortisone concentration was high, interleukin-8 concentration amounted to  $0.38 \pm 0.049$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ). These values are given on Figure 5. This observed discrepancy can be an evidence that hydrocortisone produces inhibiting effects on immune cells when an influencing factor occurs in low concentrations. We didn't reveal any significant influence exerted by hydrocortisone under high interleukin-1 concentration.

This observed discrepancy in produced effects can be explained by the following: hydrocortisone inhibits immune reaction at the initial stage in the process thus preventing the overall chain of protective reactions in a body from activating as a virus occurs in small quantities and it is not necessary to waste energy accumulated in a body.

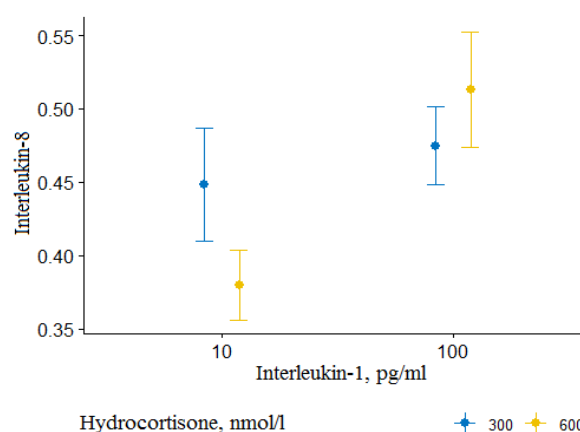


Figure 5. Average standardized interleukin-8 concentrations with error of mean under different concentration of interleukin-1 taking into account effects by hydrocortisone

Aluminum oxide exerts similar influence on interleukin-8 production under exposure to interleukin-1. Average standardized interleukin-8 concentration amounted to  $0.453 \pm 0.082$  under low interleukin-1 influence and low aluminum oxide concentration; but if aluminum oxide concentration was high, interleukin-8 concentration was equal to  $0.374 \pm 0.042$  (there was an authentic discrepancy between the obtained average values,  $p < 0.05$ ). These values are shown on Figure 6.

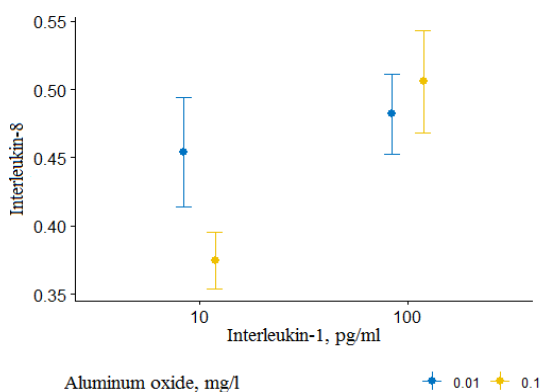


Figure 6. Average standardized interleukin-8 concentrations with error of mean under different concentration of interleukin-1 taking into account effects by aluminum oxide

We applied the least squares method (Statistica 6.0 software) to identify parameters of the model (1). The obtained equation, taking into account new quotients values, can be given as follows:

$$y(x_i) = 0,35 + x_2(0,11 - 0,13x_4 - 0,11x_5), \quad (4)$$

where  $y(x_i)$  is standardized interleukin-8 concentration, and interleukin-8 characterizes functional capabilities of immune cells;

$x_2$  is standardized interleukin-1 concentration;

$x_4$  is standardized hydrocortisone concentration;

$x_5$  is standardized aluminum oxide concentration.

The obtained relationship allows to adjust parameters of changes in infected cells quan-

tity under an immune response in the system of equations (2). The summand  $k_5 C_{NK} C_I C_{IL_2}$  in the equation (2.2) describes destruction of infected cells in a target organ by NK-cells without new viruses occurrence.

To assess what influence was produced on solutions to the system of equations by effects produced by aluminum oxide on NK-cells functional capabilities, we applied a function with an adjusted quotient  $k_5$  together with the previously obtained relationship (4). Figure 7 shows results of modeling for three different aluminum oxide concentrations.

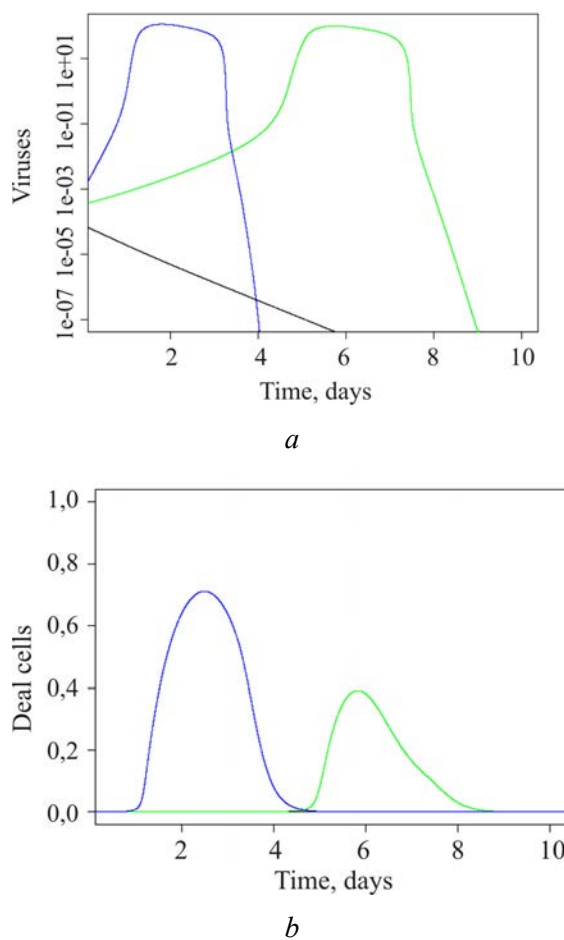


Figure 7. a) the relationship between viruses concentration in a body and time under different aluminum concentrations, b) the relationship between damage done to a target organ and time under different aluminum concentrations (the black line corresponds to aluminum oxide concentration equal to 0.01 mg/l; the green line, 0.1 mg/l; the blue line, 1 mg/l)

The obtained results revealed that when aluminum concentration was minimal (0.01 mg/l), a body recovered quickly, without any apparent symptoms of the disease. Under average exposure to aluminum (0.01 mg/l) and the same initial conditions, dynamics of changes in flu viruses quantities corresponded to a typical average clinical course of the disease. But if influence exerted by an external factor was high (1 mg/l), viruses swiftly reproduced themselves and it resulted in almost fatal damage to a target organ. There are the following gradations for damages to a target organ tissue corresponding to a clinical form of a disease: damage to less than 8-10% of a tissue corresponds to a mild disease; 10-20%, an average disease; 20-25%, a grave disease; when more than 25-30% of a target organ tissue is damaged, lethal outcome is rather probable [39].

**Conclusions.** So, we developed a predictive mathematical model that describes functioning of the regulatory systems under exposure to a virus infection; this model allows to take into account influence exerted by exposure to chemical factors on key components in

the immunity. We suggested an algorithm for conducting an experiment on identifying some parameters of influence exerted by chemical factors on interaction between the neuroendocrine and immune systems.

We examined immune regulation peculiarities via experimental research and its results revealed that there were interrelations in the system of immune evolution-controlling proteins under combined exposure to hydrocortisone (a neuroendocrine factor) and aluminum (a chemical factor). The developed mathematical model showed that detected regularities related to influences exerted by chemical and physiological factors of various genesis on functions performed by immune cells were non-linear. These detected relationships can be applied to efficiently predict disorders in an immune response and to assess interactions between the neuroendocrine and immune systems as such interactions determine adaptation reserves of a body under technogenic exposure.

**Funding.** The research was not granted any sponsor support.

**Conflict of interests.** The authors state there is no any conflict of interests.

## References

1. Heijnen C.J. Receptor regulation in neuroendocrine-immune communication: current knowledge and future perspectives. *Brain, behavior, and immunity*, 2007, vol. 21, no. 1, pp. 1–8.
2. Pace T.W., Negi L.T., Adame D.D., Cole S.P., Sivilli T.I., Brown T.D., Issa M.J., Raison C.L. Effect of compassion meditation on neuroendocrine, innate immune and behavioral responses to psychosocial stress. *Psychoneuroendocrinology*, 2009, no. 34, pp. 87–98.
3. Ashley N.T., Demas G.E. Neuroendocrine-immune circuits, phenotypes, and interactions. *Hormones and Behavior*, 2017, vol. 87, pp. 25–34.
4. Suarez E.C., Sundy J.S., Erkanli A. Depressogenic vulnerability and gender-specific patterns of neuro-immune dysregulation: What the ratio of cortisol to C-reactive protein can tell us about loss of normal regulatory control. *Brain, Behavior, and Immunity*, 2015, no. 44, pp. 137–147.
5. Lanin D.V., Zaitseva N.V., Dolgikh O.V. Neuroendokrinnye mekhanizmy regulyatsii funktsii immunnoi sistemy [Neuroendocrine Mechanisms for Regulation of Immune System]. *Uspekhi sovremennoi biologii*, 2011, no. 2, pp. 122–134 (in Russian).
6. Bellavance M., Rivest S. The neuroendocrine control of the innate immune system in health and brain diseases. *Immunological Reviews*, 2012, vol. 248, no. 1, pp. 36–55.
7. Miyake S. Mind over cytokines: Crosstalk and regulation between the neuroendocrine and immune systems. *Clinical and Experimental Neuroimmunology*, 2012, vol. 3, no. 1, pp. 1–15.
8. Poletaev A.B., Morozov S.G., Kovalev I.E. Regulyatornaya metasistema (immunoneiroendokrinnyaya regulyatsiya gomeostaza) [Regulatory Metasystem (Immunoneuroendocrine regulation of Homeostasis)]. Moscow, Meditsina Publ., 2002, 166 p. (in Russian).
9. Chapman C.R., Tuckett R.P., Song C.W. Pain and Stress in a Systems Perspective: Reciprocal Neural, Endocrine, and Immune Interactions. *Journal of Pain*, 2008, vol. 9, no. 2, pp. 122–145.

10. Savilov E.D., Mal'tsev M.V. Epidemiologicheskaya kharakteristika virusnogo gepatita S v usloviyakh krupnogo promyshlennogo goroda. *Zhurnal mikrobiologii, epidemiologii i immunobiologii*, 2007, no. 1, pp. 70–71 (in Russian).

11. Stepanenko L.A., Il'ina S.V., Savilov E.D. Osobennosti sostoyaniya spetsificheskogo immuniteta k upravlyaemym infektsiyam u detei (na primere kori i poliomielita) v usloviyakh vozdeistviya tekhnogennoi nagruzki [Features of a condition of specific immunity to controlled infections at children (on an example of measles and poliomyelitis) in conditions of technogenic influence of loading]. *Byulleten' Vostochno-Sibirskogo nauchnogo tsentra Sibirskogo otdeleniya Rossiiskoi akademii meditsinskikh nauk*, 2007, no. S3, pp. 66–68 (in Russian).

12. Stepanenko L.A., Savchenkov M.F., Il'ina S.V., Anganova E.V., Savilov E.D. Otsenka sostoyaniya immunnoi sistemy detskogo naseleniya kak markera tekhnogennoho zagryazneniya okruzhayushchei sredy [An assessment of the immune status of the children population as a marker of technogenic pollution of the environment]. *Gigiena i sanitariya*, 2016, vol. 95, no. 12, pp. 1129–1133 (in Russian).

13. Lanin D.V. Analiz koregulyatsii immunnoi i neuroendokrinnoi sistem v usloviyakh vozdeistviya faktorov riska [The analysis of the co-regulation between the immune and neuroendocrine systems under exposure to risk factors]. *Analiz riska zdorov'yu*, 2013, no. 1, pp. 73–81 (in Russian).

14. Zaitseva N.V., Shur P.Z., Mai I.V., Kir'yanov D.A. Metodicheskie podkhody k otsenke integral'nogo riska zdorov'yu naseleniya na osnove evolyutsionnykh matematicheskikh modelei [Approaches to the assessment of integrated health risk population based on evolution of mathematical models]. *Zdorov'e naseleniya i sreda obitaniya*, 2011, no. 10, pp. 6–9 (in Russian).

15. Zaitseva N.V., Trusov P.V., Shur P.Z., Kir'yanov D.A., Chigvintsev V.M., Tsinker M.Yu. Metodicheskie podkhody k otsenke riska vozdeistviya raznorodnykh faktorov sredy obitaniya na zdorov'e naseleniya na osnove evolyutsionnykh modelei [Methodical approaches to health risk assessment of heterogeneous environmental factors based on evolutionary models]. *Analiz riska zdorov'yu*, 2013, no. 1, pp. 3–11 (in Russian).

16. Zaitseva N.V., Kiryanov D.A., Lanin D.V., Chigvintsev V.M. A mathematical model of the immune and neuroendocrine systems mutual regulation under the technogenic chemical factors impact. *Computational and Mathematical Methods in Medicine*, 2014, vol. 2014 (in Russian).

17. Zabel P., Horst H.J., Kreiker C., Schlaak M. Circadian rhythm of interleukin-1 production of monocytes and the influence of endogenous and exogenous glucocorticoids in man. *Klinische Wochenschrift*, 1990, vol. 68, no. 24, pp. 1217–1221.

18. Kerdiles Y., Ugolini S., Vivier E. T cell regulation of natural killer cells. *The Journal of Experimental Medicine*, 2013, vol. 210, no. 6, pp. 1065–1068.

19. Andrew M.E., Churilla A.M., Malek T.R., Braciale V.L., Braciale T.J. Activation of virus specific CTL clones: antigen-dependent regulation of interleukin 2 receptor expression. *The Journal of Immunology*, 1985, vol. 2, no. 134, pp. 920–925.

20. Muraguchi A., Kehrl J.H., Longo D.L., Volkman D.J., Smith K.A., Fauci A.S. Interleukin 2 receptors on human B cells. Implications for the role of interleukin 2 in human B cell function. *The Journal of experimental medicine*, 1985, vol. 161, no. 1, pp. 181–97.

21. Demas G.E., Adamo S.A., French S.S. Neuroendocrine-immune crosstalk in vertebrates and invertebrates: Implications for host defence. *Functional Ecology*, 2011, vol. 25, no. 1, pp. 29–39.

22. Haus E., Smolensky M.H. Biologic rhythms in the immune system. *Chronobiology international*, 1999, vol. 16, no. 5, pp. 581–622.

23. Marchuk G.I., Petrov R.V., Romanyukha A.A., Bocharov G.A. Mathematical model of antiviral immune response. I. Data analysis, generalized picture construction and parameters evaluation for hepatitis B. *Journal of Theoretical Biology*, 1991, vol. 151, no. 1, pp. 1–40.

24. Bocharov G.A., Romanyukha A.A. Mathematical model of antiviral immune response III. Influenza A virus infection. *Journal of Theoretical Biology*, 1994, vol. 167, no. 4, pp. 323–360.

25. Joklik W.K., B.N. Fields, eds. Interferons. New York: Raven Press Publ., 1985, pp. 281–307.

26. Tamura S.I., Iwasaki T., Thompson A.H., Asanuma H., Chen Z., Suzuki Y., Aizawa C., Kurata T. Antibody-forming cells in the nasal-associated lymphoid tissue during primary influenza virus infection. *Journal of General Virology*, 1998, vol. 79, no. 2, pp. 291–299.

27. Keenan K.P., Combs J.W., McDowell E.M. Regeneration of hamster tracheal epithelium after mechanical injury. *Virchows Archiv B Cell Pathology Including Molecular Pathology*, 1983, vol. 42, no. 1, pp. 231–252.
28. G.A. Bocharov, A.A. Romanyukha. Mathematical model of antiviral immune response III. Influenza A virus infection. *Journal of Theoretical Biology*, 1994, vol. 167, no. 4, pp. 323–360.
29. Zhdanov V.M., Bukrinskaya A.G. Reproduktsiya miksovirusov (virusov grippa i skhodnykh s nimi). Moscow, Medicina Publ., 1969, 280 p. (in Russian).
30. Bergeron Y., Ouellet N., Deslauriers A., Simard M., Olivier M., Bergeron M. Cytokine kinetics and other host factors in response to pneumococcal pulmonary infection in mice. *Infection and Immunity*, 1998, vol. 66, no. 3, pp. 912–922.
31. Gloff C., Wills R., B. Ferraiolo, eds. Pharmacokinetics and Metabolism of Therapeutic Cytokines. Plenum Press Publ., New York, 1992, pp. 127–150.
32. Felig P., Frohman L., eds. Endocrinology and metabolism. New York, McGraw-Hill Publ., 2001, 1562 p.
33. B.J. Carroll, F. Cassidy, D. Naftolowitz [et al.]. Veldhuis Pathophysiology of hypercortisolism in depression. *Acta Psychiatrica Scandinavica*, 2007, vol. 115, pp. 90–103.
34. Vinther F., Andersen M., Ottesen J.T. The minimal model of the hypothalamic-pituitary-adrenal axis. *Journal of Mathematical Biology*, 2011, vol. 63, no. 4, pp. 663–690.
35. Brand J.M., Schmucker P., Breidhardt T., Kirchner H. Upregulation of IFN- $\gamma$  and Soluble Interleukin-2 Receptor Release and Altered Serum Cortisol and Prolactin Concentration during General Anesthesia. *Journal of Interferon & Cytokine Research*, 2001, vol. 10, no. 21, pp. 793–796. DOI: 10.1089/107999001753238024
36. Yoneda K., Osaki T., Yamamoto T., Ueta E. Effects of tumour necrosis factor-alpha (TNF-alpha), IL-1 beta and monocytes on lymphokine-activated killer (LAK) induction from natural killer (NK) cells and T lymphocytes. *Clinical & Experimental Immunology*, 1993, vol. 2, no. 93, pp. 229–236.
37. Callewaert D.M., Moudgil V.K., Radcliff G., Waite R. Hormone specific regulation of natural killer cells by cortisol. Direct inactivation of the cytotoxic function of cloned human NK cells without an effect on cellular proliferation. *FEBS Letters*, 1991, vol. 1, no. 285, pp. 108–110.
38. Marchuk G.I., Berbentsova E.P. Ostrye pnevmonii. Immunologiya, otsenka tyazhesti, klinika, lechenie. Moscow, Nauka Publ., 1989, 304 p. (in Russian).
39. Callewaert D.M., Moudgil V.K., Radcliff G., Waite R. Hormone specific regulation of natural killer cells by cortisol. Direct inactivation of the cytotoxic function of cloned human NK cells without an effect on cellular proliferation // *FEBS Lett*, 1991, vol. 285, no. 1, pp. 108–110.
40. Wohlfart C. Neutralization of Adenoviruses: Kinetics, Stoichiometry, and Mechanisms // *J. Immunol.* 1988, vol. 62, no. 7, pp. 2321–2328.

*Trusov P.V., Zaitseva N.V., Chigvintsev V.M. Assessing risks of adverse clinical course and outcome of an infectious disease with mathematical modeling of exposure to environmental factors on the example of aluminum oxide. Health Risk Analysis, 2019, no. 1, pp. 17–29. DOI: 10.21668/health.risk/2019.1.02.eng*

Received: 01.02.2019

Accepted: 28.02.2019

Published: 30.03.2019